

Research

Anthropophilic mosquitoes and malaria transmission in a tropical rain forest area of Nigeria

*¹Oyewole, I. O., ²Ibidapo, C. A., ³Oduola, A. O., ⁴Obansa, J. B. & ⁴Awolola, T. S

¹Babcock University, Ilishan-Remo, Department of Basic and Applied Sciences, PMB 21244, Ikeja, Lagos 100 001, Nigeria. ²Lagos State University, Department of Zoology, Ojo, Lagos, Nigeria. ³University of Lagos, Department of Zoology, Akoka, Nigeria. ⁴Institute for Medical Research, Public Health Division, Lagos, Nigeria

Received: 8 June, 2005

Revision accepted: 23 Aug., 2005

*Correspondence author <sicoprof@yahoo.com>

Abstract

Adult female *Anopheles* mosquitoes attracted to man were collected twice a month (December 2003-January 2005) from communities located in tropical rain forest region of Ikenne and Remo North Local Government Areas of Ogun State, Nigeria. The collection was made on Human volunteers from indoor and outdoor between 1900-0600h following WHO (1975) standard procedure. A total of 1500 Anopheline mosquitoes were collected and identified using both morphological and molecular techniques. This constitutes 790 *An. gambiae* (52.7%), 555 *An. arabiensis* (37%), and 155 *An. funestus* group (10.3%). The indoor catch 807(53.8%) predominated over the outdoors 693(46.2%) which constitute mainly of *An. rivulorum* and *An. arabiensis*. The biting activity observed indoor was significantly higher than outdoor ($P < 0.05$) with a ratio of 10.1: 9.60, indoor to outdoor. Although, there was no significant difference in the seasonal biting activity ($P > 0.01$), the mosquito species that fed exclusively on man were significantly higher than those with mixed blood ($P < 0.05$). The implicated malaria vectors were *An. arabiensis*, *An. gambiae* s. s. and *An. funestus* s. s. with overall infection rates of 2.3%, 2.5% and 2.9% respectively. This study emphasizes the need for accurate identification of *Anopheles* mosquitoes attracted to man in order to determine vector-host contact necessary for malaria transmission and to provide information required to formulate vector control programme.

Keywords: *Anopheles* mosquitoes, PCR identification, ELISA test, malaria, Nigeria

Introduction

Anopheline vector of malaria consists of various species with unique behaviour associated with their biting activities and transmission dynamics.

In Nigeria, the *Anopheles* mosquitoes involved in malaria transmission belong to *An. gambiae* and *An. funestus* group (Boreham et al., 1979; Molineaux & Gramiccia, 1980; Rishikesh et al., 1985). Others regarded as minor/secondary vectors include *An. moucheti* and *An. nili* (Gillies & De Meillon, 1968; Gillies and Coetzee, 1987).

The *An. gambiae sensu lato* consist of seven species, five of which are vectors of human malaria.

Amongst these are *An. gambiae* and *An. arabiensis* (the two major malaria vectors in Africa) (Gillies & De Meillon 1968), *An. merus*, *An. melas*, and *An. bwambae* (the minor vectors). The other species *An. quadriannulatus* A and B are the non-vector species from southern Africa, and Ethiopia respectively (Gillies & Coetzee, 1987; Hunt et al., 1998). The *An. funestus* group also consists of nine morphologically indistinguishable species. These include, *An. funestus sensu stricto*, *An. vaneedeni*, *An. parensis* Gillies, *An. aruni* Sobti, *An. fuscivenosus* Leesoni, *An. confusus* Evans & Leesoni, *An. leesoni* Evans, *An. rivulorum* Leesoni and *An. brucei* (Gillies & De Meillon, 1968; Gillies & Coetzee, 1987).

Studies on the biology of *Anopheles* species

in Nigeria had described *An. gambiae* s. s, *An. arabiensis*, *An. funestus*, *An. rivulorum* and *An. Leesonii* as vectors of malaria (Boreham *et al.*, 1979; Coluzzi *et al.*, 1979; Molineaux & Gramiccia, 1980; Rishikesh *et al.*, 1985; Onyabe & Conn, 2001; Awolola *et al.*, 2002). The role of anthropophilic mosquitoes in malaria transmission in southwestern Nigeria has also been documented (Awolola 2003, 2005).

The present study embarked on a longitudinal collection of man-biting mosquitoes with a view to investigating activities and habits related to malaria transmission, epidemiology and control.

Materials and Methods

Study Area

This study was carried out in Akaka, Ilara and Ijesa-Isu communities located in a forest area characterized with moderately hot and humid tropical climate in southwestern Nigeria. The two distinct seasons in this area are rainy season, which lasts from March/April to October/November, and the dry season, which lasts for the rest of the year, October/November till March/April. The mean temperature is 30°C during the dry season and 24°C in the rainy season. The mean annual rainfall is about 2000mm (Anon, 2005).

The study communities were selected based on altitude, size, house types, presence of water bodies especially slow running ones and prevailing vegetation type.

The inhabitants usually engaged in subsistence farming and petty trading for a living. The houses were constructed with either block and cement or mud with corrugated iron sheet, bamboo or palm fronds.

The study areas lack social amenities such as pipe borne water, modern health facilities and waste disposal system. The residents often keep domestic animals within their houses or nearby sheds.

Mosquito Collection and Identification

Adult female Anopheles was collected twice a month from December 2003 to January 2005. The collection was made on human volunteers stationed indoor and outdoor from 19:00-06:00 hr following the standard of WHO (1975) procedure. The samples were then preserved dry on silica gel for further analysis.

Identification of mosquito was carried out by morphological and PCR methods. The keys of Gillies and De Meillon (1968), Gillies and Coetzee (1987) were used as guides for morphological identification. Molecular assays used were the species-specific PCR techniques for confirmatory identification of the species within *An. gambiae* complex, and a cocktail PCR technique of Koekemoer *et al.* (2002) to identify the

species within *An. funestus* group.

ELISA Tests

Sporozoite ELISA

The circumsporozoite proteins of Plasmodium species present on the head and thorax of 1500 Anopheles mosquitoes were tested following the method of Wirtz *et al.* (1987). Sporozoite rates were determined photometrically at 405nm using an ELISA plate reader (Anthos 2010, Anthos Labtec GmbH, 5017 Wals/Salzburg, Austria), 30 min after the addition of substrates (Beier *et al.*, 1990). Positive ELISA reading was taken as the absorbance value greater than twice the mean of negative control.

Blood Meal ELISA

The blood meal source of 830 blood-fed samples caught outdoor and indoor was determined using the direct ELISA method of Beier *et al.* (1988).

Statistical Analysis

The relative abundance of the species was expressed as the percentage of the total number of Anopheles collected. Chi-Square tests were used to analyze differences in indoor and outdoor biting activities and sporozoite rates.

Results

A total of 1500 adult female Anopheles mosquitoes were identified by molecular assays.

The *An. gambiae* species-specific PCR identified 52.7% as *An. gambiae* s. s and 37% as *An. arabiensis* while the multiplex PCR assay identified 7% as *An. funestus* s.s and 3.3% as *An. rivulorum*.

The indoor catch 807(53.8%) predominated over the outdoors 693(46.2%). *An. gambiae* constitute the highest indoor biting species followed by *An. funestus* while *An. arabiensis* and *An. rivulorum* were mainly outdoor species (Table 1).

Overall biting activity observed indoor was significantly higher than outdoor ($P < 0.05$) with a ratio of 10.1: 9.60, indoor to outdoor. Although, outdoor biting activity was higher in the dry season during which *An. arabiensis* was the predominant species.

Table 1: Abundance of species within *An. gambiae* complex and *An. funestus* group caught indoors and outdoors

Species	Indoor collection No (%)	Outdoor collection No (%)	Total caught/ location No (%)	Indoor /Outdoor Ratio
<i>An. gambiae</i>	500(62)	290(42)	790(52.7)	1.48
<i>An. arabiensis</i>	200(24.8)	355(51.2)	555(37.0)	2.06
<i>An. funestus</i>	85(10.5)	20(2.8)	105(7.0)	3.75
<i>An. rivulorum</i>	22(2.7)	28(4)	50(3.3)	1.48
Total	807(53.8)	693(46.2)	1500	

Table 2: Seasonal variation in man-vector contact

Season	Total no collected	<i>An. gambiae</i>		<i>An. arabiensis</i>		<i>An. funestus</i>		<i>An. rivulorum</i>	
		Indoor	Outdoor	Indoor	Outdoor	Indoor	Outdoor	Indoor	Outdoor
Wet Season (April – Oct)	1013	382	105	114	164	131	13	43	60
Dry Season (Nov – March)	487	141	48	54	91	81	7	31	34
Total	1500	523	153	168	255	213	20	74	94

The wet season biting activity was more pronounced indoor and often started as early as 2000h (pre-bed time). In most of the species found indoor, biting activity lasted throughout the night from 2400-0600h (post-bed time). There was no significant difference in the seasonal biting activity ($P>0.01$). The species of *An. gambiae*, *An. funestus* were more of endophilic than exophilic as characterized by *An. arabiensis* and *An. rivulorum* irrespective of the seasonal changes (Table 2).

Blood Meal Source

The ELISA results of 830 anopheles mosquitoes that appeared blood -fed showed that 84.7% *An. gambiae* s.s, 68.8% *An. funestus* s.s, 41.6% *An. arabiensis* and 8.2% *An. rivulorum* fed on man (Table 3). The mosquito species that fed exclusively on man were significantly higher than those with mixed blood ($P<0.05$). The preference for blood meal has no relationship with the season of the year.

Table 3: Percentages indoor-outdoor blood-fed Anopheles mosquitoes and their respective hosts

Positive blood meal ELISA test							
Species	No	Indoor No (%)	Outdoor No (%)	Human No (%)	Bovine No (%)	Mixed No (%)	Other sources No (%)
<i>An. gambiae</i>	360	253 (70.2)	107 (29.8)	305 (84.7)	10 (2.8)	5 (1.4)	40 (11.1)
<i>An. arabiensis</i>	245	65 (26.5)	180 (73.5)	102 (41.06)	90 (36.7)	40 (16.4)	13 (5.3)
<i>An. funestus</i>	128	88 (68.8)	40 (31.3)	88 (68.8)	18 (14.1)	7 (5.5)	15 (11.2)
<i>An. rivulorum</i>	97	23 (23.7)	74 (76.3)	9 (8.2)	55 (56.7)	24 (15.5)	9 (9.3)
Total	830	429 (51.7)	401 (48.3)	504 (60.7%)	173 (21.0)	76 (9.2)	77 (9.3)

Sporozoite ELISA

Out of 1374 Anopheles mosquitoes examined, only 34(2.5%) tested positive for *P. falciparum* circumsporozoite antigen (CSA). The overall sporozoite rates were 10/245(2.3%) for *An. arabiensis*, 12/360(2.5%) for *An. gambiae* s. s. and 16/552(2.9%) for *An. funestus* and 0/97(0%) for *An. rivulorum* (Table 4). There was no statistical difference in the overall

sporozoite rates for *An. gambiae* s. s., *An. arabiensis* and *An. funestus* s. s. in all study sites ($P >0.05$). Although the overall sporozoite rates dropped from 2.5% in the wet season to 1.6% in the dry season.

The major malaria vectors implicated in malaria transmission in the study areas are *An. gambiae*, *An. arabiensis* and *An. funestus*.

Table 4: Percentages indoor-outdoor catches of Anopheles mosquitoes infected with Plasmodium falciparum

Species	Total no examined		Total no positive		Total n (%)
	Indoor	Outdoor	Indoor n (%)	Outdoor n (%)	
<i>An. gambiae</i> s.s	267	213	8 (3.0)	4 (1.9)	12 (2.5)
<i>An. arabiensis</i>	100	145	2 (2.0)	4 (2.8)	6(2.3)
<i>An. funestus</i>	370	182	10 (2.7)	6 (3.3)	16 (2.9)
<i>An. rivulorum</i>	23	74	0	0	0
Total	429	401	22 (5.1)	16 (3.9)	34 (2.5)

Discussion

The abundance of mosquito is closely related to the prevalence of malaria most especially in the endemic area. The mosquitoes attracted to man are the major vectors implicated in the transmission of malaria. Out of the total amount of mosquitoes collected in this study, Anopheline species account for about 60% ind-

cating a high density. Previous studies have also shown that the density of Anopheles mosquitoes is usually very high in the tropical rainforest perhaps due to the environmental friendly conditions (Awolola *et al.*, 2003).

The correct identification of the species within the taxa is very important in recognizing those

that actually fed on man with vectorial capacity to transmit malaria parasite. In this study we used molecular techniques to complement morphological identification of Anopheline species caught feeding on man. Molecular assays have been of tremendous help in solving the problem of misidentification of species especially those that are non-vectors and those that occur in sympatry but with different behaviour (Scott *et al.*, 1993; Koekemoer *et al.*, 2002, Weeto *et al.*, 2004).

Two of the members within *An. gambiae* complex which are the major vectors of malaria and which occur sympatric were *An. gambiae* s. s. and *An. arabiensis*. Our findings showed that both *An. gambiae* and *An. arabiensis* were often found in the same vicinity, although more of *An. gambiae* were found feeding on man indoor than outdoor. The biting activities of these species have been found to complement each other such that while one is endophagic, the other is exophagic (Gillies & Coetzee 1987). The population of *An. arabiensis* was significantly high compared to other species in our dry season collections. *An. arabiensis* had been reported to persist even during the dry season while *An. gambiae* were more abundant during the wet season (Rishikesh *et al.*, 1985; Awolola *et al.*, 2002). This study also showed that biting activity of Anopheline species especially those that fed indoor continues through the day and night and all year round. Although *An. arabiensis* has been described as opportunistic but largely more of zoophilic behaviour (White, 1974; Gillies & Coetzee, 1987). Other findings showed that *An. arabiensis* readily fed on both humans and cattle (Gillies & De Meillon 1968; White, 1974). This was also confirmed in this study as we found mixed blood meal in *An. arabiensis*. Species within the *An. funestus* group occurred in sympatry, but, *An. funestus* s. s. was more endophilic and anthropophilic than *An. rivulorum*. Earlier findings also showed that some members of *An. funestus* group were found in sympatry in West Africa but of different feeding behaviour (Gillies & Coetzee, 1987; Kamau *et al.*, 2003).

The present study showed that sporozoite rates in *An. gambiae* s. s., *An. arabiensis* and *An. funestus* s. s. were statistically insignificant ($P > 0.05$). This was an indication that malaria transmission was not an exclusive role of the endophilic mosquitoes. The exophilic species also play specific role in the transmission process though they showed less anthropophilic behaviour (Haridi, 1972; Ameneshewa & Service, 1996; Fettes *et al.*, 2004). The 0% sporozoite rate recorded for *An. rivulorum* showed that it is not a vector of malaria despite the fact that it fed on man. Previous findings had also showed that *An. rivulorum*

not a vector of malaria despite the fact that it fed on man. Previous findings had also showed that *An. rivulorum* and *An. lesoni* played no tangible role in malaria transmission in West Africa (Costantini *et al.*, 1999).

The knowledge of the biology and behaviour of the Anopheline species attracted to man is vital to control and prevention of malaria. This study has made scientific attempt to provide information on the anthropophilic mosquitoes and their malaria epidemiology which is pivotal to the formulation of effective control programmes.

Acknowledgements

We gratefully acknowledged the efforts of our field assistants during the field activities. This study was partly supported by the grant MIM project A30026 through the UNICEF/UNDP/World Bank/WHO Special Programme for Research and Training in Tropical Diseases (TDR) awarded to T. S. Awolola.

Reference

- Anon 2005. Nigeria physical setting. Online Nigeria Portal www.onlinenigeria.com
- Ameneshewa, B., Service, M.W. 1986. Resting habits of *Anopheles arabiensis* in the Awash River valley of Ethiopia. *Annals of Tropical Medicine and Parasitology* **90**: 515-521.
- Awolola, T. S., Okwa, O., Hunt, R. H., Ogunrinade, A. F & Coetzee, M. 2002. Dynamics of the malaria-vector populations in coastal Lagos, southwestern Nigeria. *Annals of Tropical Medicine & Parasitology* **6** (1): 75-82.
- Awolola, T. S., Ibrahim, K., Okorie, T., Koekemoer, L. L., Hunt, R. H. & Coetzee, M. 2003. Species composition and biting activities of anthropophilic Anopheles mosquitoes and their role in malaria transmission in a holo-endemic area of southwestern Nigeria. *African Entomology* **11**(2): 227-232.
- Awolola, T. S., Oyewole, I. O., Koekemoer, L. L., & Coetzee, M. 2005. Identification of three members of *Anopheles funestus* (Diptera: Culicidae) group and their role in malaria transmission in two ecological zones in Nigeria. *Royal Society of Tropical Medicine and Hygiene* **99**: 525-531.
- Beier, J. C., Perkins, P. V., Wirtz, R. A., Koros, J., Diggs, D., Gargan, T. P., Koech, D. K. 1988. Blood meal identification by direct enzyme-linked immunosorbent assay (ELISA), tested on *Anopheles* (Diptera: Culicidae) in Kenya. *Journal of Medical Entomology* **25**: 9-16.
- Beier, J. C., Perkins, P. V., Koros, J. K., Onyango, F. K., Gargan, T. P., Wirtz, R. A., Koech, D. K., Robert, C. R. 1990. Malaria sporozoite detection by dissection and ELISA to assess infectivity of Afrotropical Anopheles (Diptera: Culicidae). *Journal of Medical Entomology* **27**: 377-384.
- Boreham, P. E., Lenahan, J. K. Boulzaquet, R., Storey, J., Ashkar, T. S., Nambiar, R. & Matsushima, T.

1979. Studies on multiple feeding by *Anopheles gambiae* s. l. in a Sudan savannah area of northern Nigeria. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **73**: 418-423.
- Coluzzi, M., Sebatini, A., Petrarca, V. & Dideco, M. A. 1979. Chromosomal differentiation and adaptation to human environments in the *Anopheles gambiae* complex. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **73**: 483-497.
- Costantini, C., Sagnon, N. F., Sanogo, E., Coluzzi, M., Boccolini, D. 1999. Chromosomal and bionomic heterogeneities suggest incipient speciation in *Anopheles funestus* from Burkina Faso. *Parasitologia* **41**: 595-611.
- Fettene, M., Hunt, R. H., Coetzee, M. & Tennessee, F. 2004. Behaviour of *Anopheles arabiensis* and *An. quadrimaculatus* sp. B mosquitoes and malaria transmission in southwestern Ethiopia. *African Entomology* **12** (1): 83-87.
- Gillies, M. T. & Coetzee, M. 1987. A supplement to the Anophelinae of Africa South of the Sahara (Afrotropical Region). *Publications of the South African Institute for Medical Research* No. 55.
- Gillies, M. T. & De Meillon, B. 1968. The Anophelinae of Africa south of the Sahara (Ethiopian Zoogeographical Region). *Publications of the South African Institute for Medical Research* No. 54.
- Haridi, A. M. 1972. Partial exophily of *Anopheles gambiae* species B in the Khasem Elgirba area in eastern Sudan. *Bulletin of the World Health Organization* **46**: 39-46.
- Hunt, R. H., Coetzee, M. & Fettene, M. 1998. The *Anopheles gambiae* complex: A new species from Ethiopia. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **92**: 231-235.
- Kamau, L., Munyekenye, G. O., Koekemoer, L. L., Hunt, R. H. & Coetzee, M. 2003. A survey of the *Anopheles funestus* (Diptera: Culicidae) group of mosquitoes from ten sites in Kenya with special emphasis on population genetic structure based on chromosomal inversion Karyotypes. *Journal of Medical Entomology* **4**: 64-671.
- Koekemoer, L. L., Kamau, L., Hunt, R. H. & Coetzee, M. 2002. A cocktail polymerase chain reaction (PCR) assay to identify members of the *Anopheles funestus* (Diptera: Culicidae) group. *American Journal of Tropical Medicine and Hygiene* **66**: 804-811.
- Molineaux, L. & Gramiccia, G. 1980. The Garki project. *Research on the Epidemiology and Control of Malaria in the Sudan Savannah of West Africa*. World health Organization, Geneva.
- Onyabe, D. Y. & Conn, J. E. 2001. Population genetic structure of the malaria mosquito *Anopheles arabiensis* across Nigeria suggests range expansion. *Molecular Ecology* **10**: 2577-2591.
- Rishikesh, N., Di Deco, M. A., Petrarca, V. & Coluzzi, M. 1985. Seasonal variation in indoor resting *Anopheles gambiae* and *Anopheles arabiensis* in Kaduna, Nigeria. *Acta Tropica* **42**: 165-170.
- Service, M. W. 1977. A critical review of procedures for sampling of adult mosquitoes. *Bulletin of Entomological Research* **67**: 343-382.
- Scott, J. A., Brogdon, W. G. & Collins, F. H. 1993. Identification of single specimen of the *Anopheles* complex by polymerase chain reaction. *American Journal of Tropical Medicine and Hygiene* **49**: 520-529.
- Weeto, M. M., Koekemoer, L. L., Kamau, L., Hunt, R. H. & Coetzee, M. 2004. Evaluation of species PCR assay for the *Anopheles* group from eleven African countries and Madagascar. *Transaction of the Royal Society for Tropical Medicine and Hygiene* **98**: 142-147.
- Wirtz, R. A., Zavala, F., Charoenvit, Y., Campbell, G. H., Burkot, T. R., Schneider, I., Esser, K. M., Beaudoin, R. I., & Andre, R. G. 1987. Comparative testing of monoclonal antibodies against *Plasmodium falciparum* sporozoite for ELISA development. *Bulletin of the World Health Organization* **65**: 39-45.
- World Health Organization 1975. *Manual of Practical Entomology on Malaria*. Part 11. WHO, Geneva.
- White, G. B. 1974. *Anopheles* complex and disease transmission in Africa. *Transaction of the Royal Society of Tropical medicine and Hygiene* **59**: 291-292.